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SYNTHESIS, ANTIMICROBIAL EVALUATION AND MOLECULAR DOCKING OF SOME PYRIDINE, PYRAN, PYRAZOLINE AND/OR ISOXAZOLINE-9-(*P*-SUBSTITUTEDANILINO)-ACRIDINE DERIVATIVES

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ABSTRACT

A new series of nitrogen and/or oxygen heterocyclic- 9-(*p*-substituted anilino) acridines were synthesized for the purpose of antimicrobial evaluation. These heterocycles include pyridinecarbonitrile, iminopyridine, pyrans, acridine-chalcones, 4,5-dihydropyrazoline and/ or isoxazoline ring systems. Some representative examples of these compounds compared with other known derivatives, showed considerable antimicrobial activity against G +ve, G -ve bacteria and fungi. Molecular docking studies of the affinity of binding of these compounds in the DNA gate of the bacterial gyrase enzyme were carried out.

KEY WORDS: Acridines; Pyridines; Pyran; Pyrazolines; Isoxazolines, Antimicrobial, Molecular docking.

INTRODUCTION

Acridine derivatives are widely used in medicine as pharmaceutical agents [1-6]. Most of the acridine derivatives are focused on the 9-substituted amino or substituted anilino acridine scaffold, paying more attention on the substituents effects in the acridine and aniline rings on the DNA binding ability and cytotoxicity [7-12]. Acridines are firstly used as antibacterial and antimalarial agents with planar aromatic structures that are capable to intercalate into DNA base pairs [5,6,13]. Also acridine compounds have a long history as anticancer [1,2], anti-oxidant [3] and anti-inflammatory [4]. Their potentials as antitumor drugs have been previously examined. For example, *N,N*-dimethyl-*N'*-(1-nitro-9-acridinyl)-1,3-propanediamine (Fig. 1 Nitracrine) showed potent antineoplastic activity [14,15]. Also, 1-[(6-chloro-2-(methoxy-9-acridinyl)-amino)-3-diethylamino-2-propanol dihydrochloride (Fig. 1 Acranil) is a known antiviral agent. It exhibits interferon induction and radio protective

activities [16]. It is generally assumed that most of the biological activity of these compounds is connected with their well-established DNA intercalation due to its flat structure. Each intercalating drug binds strongly to particular base pairs as result of several interactions, ranging from van der Waals forces to the formation of hydrogen bonds with adjacent nucleobases [17-19]. Their binding to DNA is dependent on a number of structural factors, particularly the substituents on both the acridine nucleus and the 9- amino or anilino function groups [20-24]. 9-(pyridin-2'-yl)-aminoacridines were prepared and analysed for their ability to change the thermal denaturation temperature of genomic calf thymus DNA [25]. On the other hand, several pyridones, pyrans, pyrazolines and/or isoxazoline derivatives were found to possess considerable antimicrobial activity [26-31].

The aim of the present work is to synthesize a new series of acridine derivatives incorporated at its 9-position

into the above mentioned heterocyclic systems through a *p*-iminophenyl moiety. The presence of this binary system in one molecule may results in a dual mode of biological action which in turn may increase the antimicrobial activity.

Chemistry

The chemical modification of 9-anilinoacridines, such as the introduction of different substitutions or heterocyclic ring systems were allowed expansion of SAR studies to afford new insight into molecular interaction at receptor level in the parasite cell. Recently, it is well established that slight structural modification on the substitution at the 9- position of acridine ring may bring various biological activities [25,32].

Also, it is well known that chalcones are considered as excellent starting materials for the synthesis of 2(1*H*) pyridones in a reaction where Aldol condensation product reacts with ethyl cyano acetate in the presence of ammonium acetate [22]. This procedure is time consuming and experiences some difficulties especially with propenones bearing heterocyclic moieties. So, in the present work, a method for one-step synthesis of some new cyanopyridones, cyanoiminopyridines and aminopyrans incorporated to acridine through a *p*-iminophenyl group at the 9-position of the acridine moiety is reported [26] by which good yields of the products were obtained.

The starting material of choice was 9-(*p*-acetylanilino)-acridine (**1**) [33] which allowed to react with ethyl cyano acetate and 3,4-dihydroxybenzaldehyde in the presence of ammonium acetate to afford the corresponding 3-cyano-2(1*H*)pyridone derivative **2e** as an example of the one step Michael condensation reaction according to a reported method [26] (Scheme 1).

The IR spectra of compounds **2a-e** showed bands due to NH (3350), typical 2(1*H*) pyridone NH (3160), -CN (2200) cm⁻¹ and C=O (1660). ¹H NMR spectrum of the new compound **2e** (DMSO-*d*₆) showed signals at δ 7.2-8.2 ppm (m, 16H aromatic), δ 8.3 (s, 1H amide) and at δ 8.7 (s, 1H, NH). MS spectrum of **2e** showed a peak at *m/z* 194.1 (C₁₃H₉N₂) (100% base peak) and M⁺ at *m/z* 496 C₃₁H₂₀N₄O₃ (9%).

In the same manner, reaction of **1** with malononitrile and the appropriate aldehyde under the same reaction conditions, afforded the corresponding 2 (1*H*) iminopyridines **3a-c**. [26]. The IR spectrum of **3b** showed bands attributable to 2-NH₂-pyridine at 3480, 3290 cm⁻¹, NH at 3220 cm⁻¹, 2180 (CN) and at 1600 cm⁻¹ (C=C). ¹H NMR of the new compound **3b** (DMSO-*d*₆) showed: δ 2.5 ppm (s, 3H, CH₃), δ 7.0-8.4 ppm (m, 17H aromatic), δ 8.8, 9.0 (s, 1H, NH and br s, 2H, NH₂) and its MS showed molecular ion peak of C₃₂H₂₃N₅ M⁺ at *m/z* 477 (5%) However, the reaction of **1** with malononitrile and an aldehyde in presence of piperidine afforded the corresponding 2-aminopyrans **4a,b** respectively (Scheme 1). The IR spectrum of **4** showed bands due to NH₂ at 3450, 3300 cm⁻¹, CN at 2115 cm⁻¹ and C=C at 1605 cm⁻¹. The MS

of **4b** showed the molecular ion peak C₃₂H₂₃N₄O M⁺ at *m/z* 479 (10%).

On the other hand, since chalcones (α,β -unsaturated ketones) are useful starting materials for the synthesis of many heterocycles such as pyrazoles, isoxazoles and pyridines, it was of interest to synthesize a new series of 9-(4-substituted anilino) acridines incorporated at the 4-position of aniline with these mentioned heterocycles for biological evaluation. Claisen-Schmidt condensation of the acetyl derivative **1** with the appropriate aldehyde in 5-10% ethanolic sodium hydroxide solution gave a poor yield of the chalcones **5** in addition to many other by-products. Also, when the reaction of **1** with aromatic aldehydes was carried out in acetic acid in presence of sodium acetate, low yields of **5** were obtained.

Another attempt was carried out by using the method described by Scholtz [34,35] in which the α,β -unsaturated ketone was prepared firstly from an aldehyde and *p*-aminoacetophenone then reacting the obtained 4-substituted cinnamoylaniline HCl salts with the required substrate, namely 9-chloroacridine. Good yields (55-70%) of the desired chalcones namely, acridine-9-[4-(3-substituted arylacryloyl)phenyl]-amines (**5a-d**) were obtained (Scheme 2). IR of **5a,b** showed band at 3400 (NH), 1668 (C=O). ¹H NMR of **5b** (DMSO-*d*₆) showed bands at δ 3.84 ppm (s, 3H, OCH₃), δ 7 (2H, dd, CH=CHCO) and at δ 7.3-8.3 (m, 16H aromatic). The MS of the new chalcone derivative **5d** displayed the molecular ion peak of C₂₆H₁₈N₂O₂ at M⁺ *m/z* 390 (10%).

The α,β -unsaturated ketones **5** were allowed to react with 98% hydrazine hydrate in ethanol to give the corresponding pyrazolines **6a,b**. When the reaction was carried out in acetic acid, the *N*-acetylpyrazoline derivative **6c** was obtained. Reaction of compounds **5a, 5d** with hydroxylamine hydrochloride in ethanolic sodium hydroxide solution afforded the corresponding isoxazolines **7a,b** respectively. Structures of the pyrazolines **6** and the isoxazolines **7** were inferred from the chemical analyses and spectral data. IR spectrum of **6c** showed absorption bands at 3280 cm⁻¹ (NH), 1668 cm⁻¹ (C=O, acetyl), and at 1600 cm⁻¹ (C=C). ¹H NMR of **6c** (DMSO-*d*₆) showed bands at δ 2.4 (s, 3H, acetyl), 3.4, 4.9 (dd, dd, 1H, 1H, -CH₂ of pyrazoline), 4.2 (dd, 1H, CH of pyrazoline), 7.2-8.3 (m, 17H aromatic) and at 9.3 (s, 1H, NH). MS of **7a** showed peaks of M⁺ C₂₈H₂₁N₃O at *m/z* 415.2 (1%) and C₁₃H₉N₂ at *m/z* 194 (100%), and its IR spectrum showed bands at 3310 (NH), 3075 (CH aromatic), 2940 (CH-aliphatic), and 1600 (C=C). Reaction of **5a** with ethyl cyano acetate in the presence of ammonium acetate at 150°C, afforded the same 1,6-dihydro-3-cyano-2(1*H*) pyridone derivative **2a** that previously obtained by the one step reaction (Scheme 1) as indicated by its similar melting and mixed melting point (195-7°C) but in 30% yield compared to the one pot reaction (66% yield) (Scheme 2).

EXPERIMENTAL

Melting points are uncorrected and were taken an electrothermal capillary melting point apparatus the IR spectra were recorded (KBr disks) on Perkin Elmer model 137 infracord spectrophotometer the ¹H NMR spectra were measured in DMSO-*d*₆ on Joel Ex-27 MHz spectrometer. The mass spectra were recorded on GCMS-Q 1000 Ex Shimadzu gas chromatography MS apparatus Microanalyses were carried out at the microanalytical unit, Faculty of Science, Cairo University and National Research Centre, Egypt. Carbon, Hydrogen and Nitrogen analyses of the new compounds were found to be identical with the calculated molecular weights within a range or error between (+ or - 0.4%).

6-[4-(Acridin-9-ylamino)phenyl]-2-oxo-4-(3,4-dihydroxyphenyl)-1,2-dihydropyridine-3-carbonitrile (2e) General method [26].

A mixture of the acetyl compound **1**(24) (3.129, 0.01 mole), ethylcyanoacetate (1.13g, 0.01 mol), 3,4-dihydroxybenzaldehyde (0.01 mol) and ammonium acetate (4.62g, 0.06 mol) in 50 ml n-butanol was heated under reflux for 6 hrs. A crystalline solid was obtained, filtered off, washed with water then with cold ethanol and crystallized from ethanol to give compound **2e**, m.p. 200 °C(ethanol) in 65% yield, Anal, C₃₁H₂₀N₄O₃, Calcd. C 75.00, H 4.03, N 11.29, Found: C 74.60, H 3.85, N 11.41%. Melting points of compounds 2a-d are 197,255,220 and 210 °C respectively [26].

6-[4-(Acridin-9-ylamino)phenyl]-2-imino-4-aryl-1,2-dihydropyridine-3-carbonitriles (3a-c):General method [26].

A mixture of **1** (3.12 gm, 0.01 mol) malononitrile (0.01 mol), p-tolualdehyde (0.01 mol) and ammonium acetate (0.985g; 0.0128 mol) in n-butanol was refluxed for 3 hours. The solid formed was filtered, washed with H₂O then petroleum ether, dried and crystallized from ethanol to give compound **3b**, m.p 198-200 °C (ethanol), Anal, C₃₂H₂₃N₅, Calcd C, 80.50, H 4.82, N, 14.67, Found, C, 80.72, H, 5.15, N, 14.90%. Melting points of compounds 3a and 3c are 270 and 278 °C respectively [26].

6-[4-(Acridin-9-ylamino)phenyl]-2-amino-4-aryl-4H-pyran-3-carbonitriles (4a,b):

A mixture of **1** (3.12g; 0.01 mol), malononitrile (0.65g, 0.01 mol), an appropriate aldehyde (0.01 mol) and few drops of piperidine in n-butanol (50 ml) was refluxed for 5 hours. The solid formed was filtered, washed with H₂O then cold ethanol and crystallized from dioxane to give compounds **4a,b**, m.p. 280, 292 °C respectively

1-[p-(Acridin-9-ylamino)phenyl]-3-aryl-propen-1-ones (5a-d):

General method:

A mixture of 9-chloroacridine [33] (2g, 0.01 mol)

and the appropriate substituted cinnamoyl aniline hydrochlorides (0.01 mol), prepared by the method of Scholtz [34], in ethanol (20 ml) and few drops piperidine was refluxed for 4h. The reaction mixture was cooled and the precipitated material was filtered off, crystallized from the proper solvent to give compounds **5a-d**. 1-[p-(Acridin-9-ylamino)phenyl]-3-(furan-2-yl)-propen-1-one (**5d**), m.p. 260 °C (AcOEt), Analysis, C₂₆H₁₈N₂O₂, (390) Calcd. C, 80.02, H, 4.61, Found, C, 79.55, H, 4.90%

9-[p-(5-Aryl-4,5-dihydro-1H-pyrazolin-3-yl) anilino] acridines (6a-c).

General method:

A mixture of the chalcone derivatives **5a,b** (0.015 mol) and 98% hydrazine hydrate (0.5g, 0.015 mol) in absolute ethanol (10 ml) was refluxed for 3h. The precipitated product was filtered off and crystallized from the proper solvent to give compounds **6a,b** respectively. When a mixture of compound **5a** (6g, 0.015 mol) and 98% hydrazine hydrate (0.5g, 0.015 mol) in glacial acetic acid (5 ml) was refluxed for 4h a precipitated product of N-acetylpyrazoline derivative **6c** was filtered off and crystallized from acetone.

Compound 6a, m.p. 212-213 °C (ethanol), Analysis, C₂₈H₂₂N₄, (414) , Calcd., C, 81.15, H, 5.31, N, 13.52, Found, 80.90, H, 5.70, N, 14.02% **Compound 6b**, m.p. 196-7 °C (ethanol), Analysis, C₂₉H₂₄N₄O (444), Calcd., C, 74.17, H, 5.40, N,12.61, Found, C,73.90, H, 5.70, N, 12.40%. **Compound 6c**, m.p. 186-7 °C (acetone), Analysis, C₃₀H₂₄N₄O (456), Calcd., C, 78.84, H, 5.26, N12.28, Found, C, 78.80, H, 5.46, N, 12.52%.

9-[p-(5-Aryl-4,5-dihydro-1H-isoxazolin-3-yl) anilino] acridines (7a,b).

A mixture of compound **5a,d** (0.015 mol) hydroxylamine hydrochloride (1 g, 0.015 mol) and sodium hydroxide (0.1g) in absolute ethanol (5 ml) was refluxed for 8 hours, and then poured onto ice water. The obtained precipitate was filtered off and crystallized from ethanol to give compounds **7a,b**. respectively. **Compound 7a**: m.p. 214 °C (ethanol), Analysis, C₂₈H₂₁N₃O, (415), Calcd. C, 80.69, H, 5.06, N, 10.12, Found, C 81.08, H, 4.95, N, 10.10.41%. **Compound 7b**: m.p. 200-201 °C (ethanol), Analysis, C₂₆H₁₉N₃O₃ (405), Calcd. C, 77.03, H, 4.69, N, 10.37, Found, C, 76.85, H, 4.88, N, 10.50%

6-[4-(Acridin-9-ylamino)phenyl]-2-oxo-4-phenyl-,2-dihydro-pyridine-3-carbonitrile (2a):

Method 2: From the chalcone 5a

A mixture of compound **5a** (0.4g, 0.001 mol), ethylcyanoacetate (0.226g, 0.002 mol) and ammonium acetate (0.308g, 0.004 mol) in n-butanol (5 ml) was heated under reflux for 5h. A crystalline solid was separated, filtered and crystallized from ethanol to give compound **3a** in 30% yield. The obtained product was found to be

identical with that obtained by the one pot reaction from compound **1** (Scheme 1). M.p. 196 °C mixed m.p. 194°C.

Biology

The selected compounds were tested against *Escherichia Coli*, *Pseudomonas Aeruginosa*, *Staphylococcus Aureus*, *Sarcina Lutea*, *Bacillus Subtilis*, *Mycobacterium Phlei*, and *Candida Albicans* and *Aspergillus Niger*. The antimicrobial activity of the tested compounds was determined in side-by-side with Nalidixic acid [36] as antibacterial reference drug or Clotrimazol [37] as antifungal reference drug.

MATERIALS

All microorganisms used were obtained from the culture collection of the department of Microbiology and Immunology, Faculty of Pharmacy, Helwan University. The compounds were tested against *Escherichia Coli*, *Pseudomonas Aeruginosa*, and *Staphylococcus Aureus* in nutrient both, pH 7.0 and against *Bacillus Subtilis* in lacto brain heart infusion broth and against *Sarcin Lutea*, and *Candida Albicans* in broth containing 1% neopeptone, 2% dextrose with pH 5.7. Media for disc sensitivity tests were nutrient agar and Muller-Hinton agar (MHA) purchased from Difco. The disc diameter was 5mm. Non sterile powder of tested compounds were dissolved in DMSO to yield 5,000 ug/ml passed through 0.0002mm membrane filters (Millipore corp. Bedford, Mass).

Disc diffusion test

20 ml of Muller-Hinton agar (MHA) at 55°C, inoculated with 1 ml of the-microbial culture (10^6 CFU/ml), was poured in sterile Petri dish and left to solidify. A sterile filter paper disc impregnated with solution of the compound under testing (0.1 mg/ ml in DMF) was placed on the surface of agar, and the plate was incubated over night at 37°C. The diameter of the zone of inhibition was measured and compared with the standard zone produced by Nalidixic acid only for antibacterial evaluation and with Clotrimazol for antifungal evaluation [38].

Sequence alignment

Alignment was done for the fasta sequence for all *Bacillus subtilis* (Uniprot code = P39814) and *E-coli* (uniprot code = P06612) and *Staph. aerus* (uniprot code = Q2FHI8) using clustal omega program

Molecular docking

Docking was done using molecular operating environment [39,40].

RESULT AND DISCUSSION

Table 1 show:

- Compounds **2a-e** and **6a,d** showed high significant activity against *Bacillus Subtilis*, compound **1** is less active,

and compound **2b** is moderately active against *Staphylococcus Aureus*.

- Compound **1** only showed moderate activity against *Eschericia Coli*, compared with Nalidixic acid.
- Compound **2e** showed moderate activity against *Mycobacterium Pheli*.
- Compound **6a** showed moderate activity against *Aspergillus Niger* (fungus) compared with the highly active Clotrimazole. 5- All compounds are not active against *Sarcina Lutea*, *Candida Albicans*, *Pseudomonas Aergmosa*.

Sequence alignment:

The antimicrobial activity of the compounds showed that all compounds were active on the *Bacillus Subtilis* and some of them were active against both *E. Coli* and *S. Aureus*. In order to interpret this sequence alignment between the gyrase enzyme of *Bacillus Subtilis* against both *E. Coli* and *S. Aureus* was performed which revealed incomplete identity between them as shown in Fig 2 and 3.

Molecular docking results:

Molecular docking was done to predict all possible orientations of the tested compounds and to find out a possible explanation of the activity of all compounds against *Bacillus Subtilis*. According to the literature it was reported that acridine derivatives have a high affinity of binding in the DNA gate of bacterial gyrase [41-43].

DNA gate has a catalytic Tyrosine residues (Tyr 123) that acts as nucleophiles by their hydroxyl groups to attack the DNA and participate in the DNA cleavage process. In the catalytic area of DNA gate there are also some other residues that interacted together for the integrity of the DNA binding site such as; Ala 68, Gly 72, Gly 76, Tyr 150 and Arg 69. These residues enable the gate to form a close as well.

The ability of any compound to interact with the Tyr 123 (the main catalytic residue) will inhibit the DNA binding. Also, the blocking of DNA gate by planar structures like acridines may help in that as well.

According to the docking results it was observed that compound **1** interacted by a hydrogen bond that is formed between its C=N and -OH of Tyr 123. Compound **2A** had two modes of binding; one by a hydrophobic interactions with Arg 122 and the second by hydrogen bond with Tyr 123. The same was observed with compounds **2B** and **2C**. Compound **2D** participated by its nitrile group to form a hydrogen bond with Tyr 123. Compound **5c** had hydrophobic interactions due to the $\pi - \pi$ interactions formed by its p-chlorophenyl ring. Compound **6A** showed a hydrogen bond by its pyrazole -NH with Arg 122.

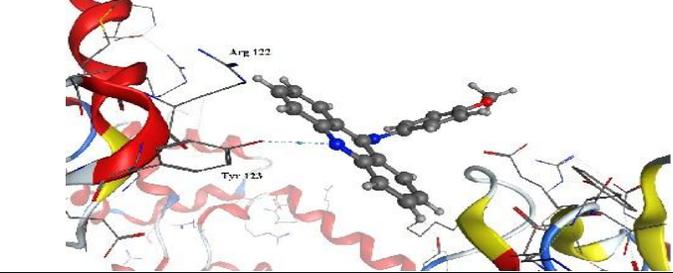
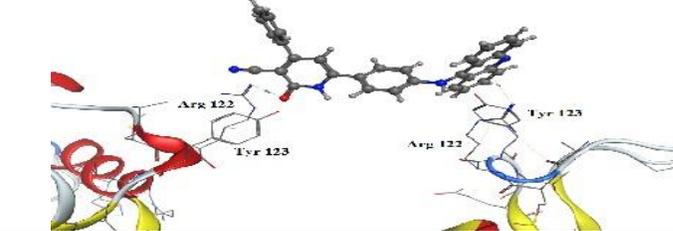
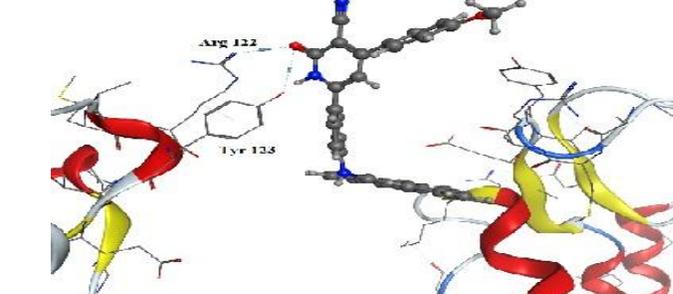
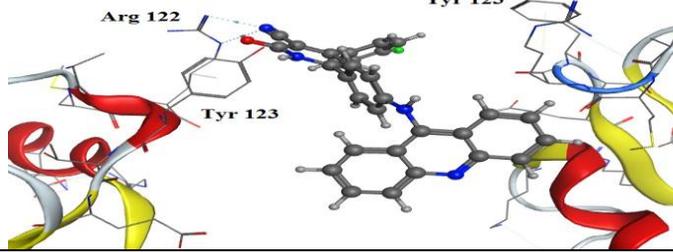
It is also predicted that all compounds have nitrile group may interact by a covalent bond with the Tyr nucleophilic center (Table 2 shows all docking results).

Table 1. The antimicrobial screening of some new acridine compounds

Comp. No.	<i>B. Auliblis</i>	<i>E. Coli</i>	<i>S. Aureus</i>	<i>S.Lutea</i>	<i>P.Aeruginosa</i>	<i>M. Plieli</i>	<i>C. Albican</i>	<i>A. Niger</i>
1	+++	+++	-	-	-	.		-
2a	+++++		-	-	-	-	-	-
2b	+++++	-	++++	-	-	-	-	-
2c	+++++	-	-	-	-	-	-	-
2d	+++++	-	+	-	-	-	-	-
2e	+++++	-	-	-	-	++++	-	-
5c	++++	-	-	-	-	-	-	-
6a	+++++	+	-	-	-	-	-	++++
NA	+++++	+++++	+++++	+++	+	++	-	(-)
Clotrimazole	(-)	(-)	(-)	(-)	(-)	(-)	(-)	(+++++)

+++++ = highly significant +++++ = moderately significant
 +++ = slight significant ++ = fairly significant
 + = weakly significant - = inactive (-) = not test.

Table 2. Docking of all tested compounds against Bacillus subtilis gyrase

		Affinity Kcal/mol	Residues
1		-11.84	Arg 122 and Tyr 123
2A		-12.35	Arg 122
2B		-12.25	Arg 122 and Tyr 123
2C		-12.40	Arg 122 and Tyr 123

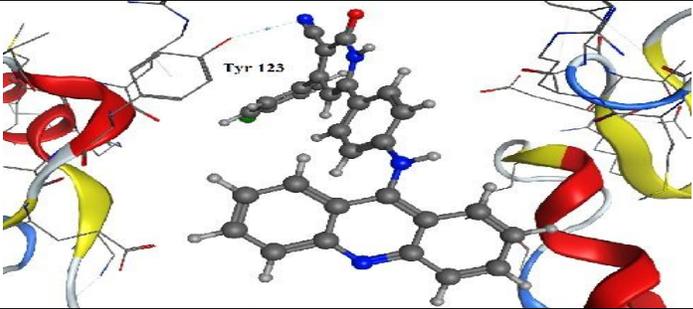
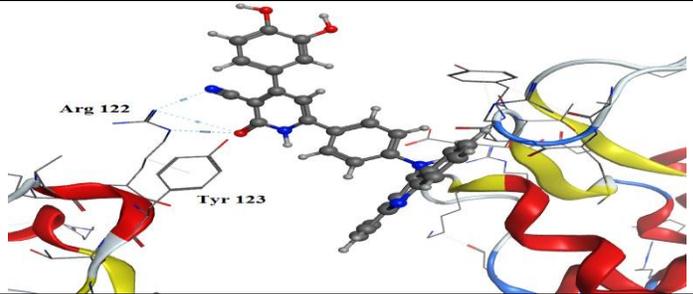
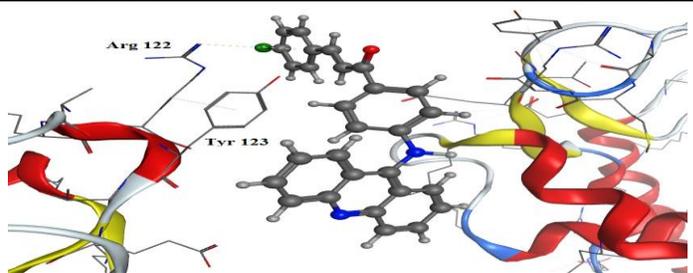
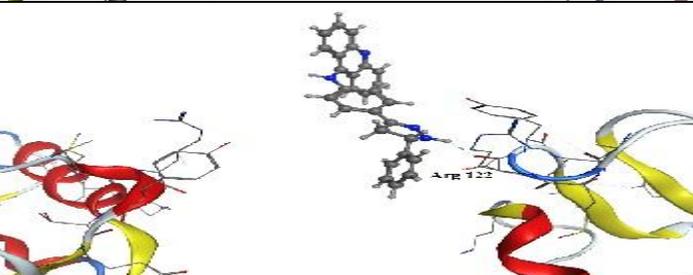
2D		-11.95	Arg 122 and Tyr 123
2E		-12.33	Arg 122 and Tyr 123
5C		-12.12	Arg 122 and Tyr 123
6A		-11.75	Arg 122

Figure 1. Chemical structures: 1, Nitracrine, 2, Acridine

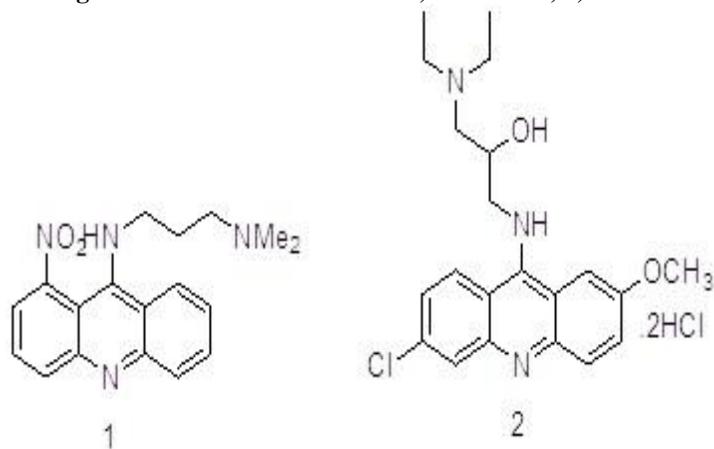


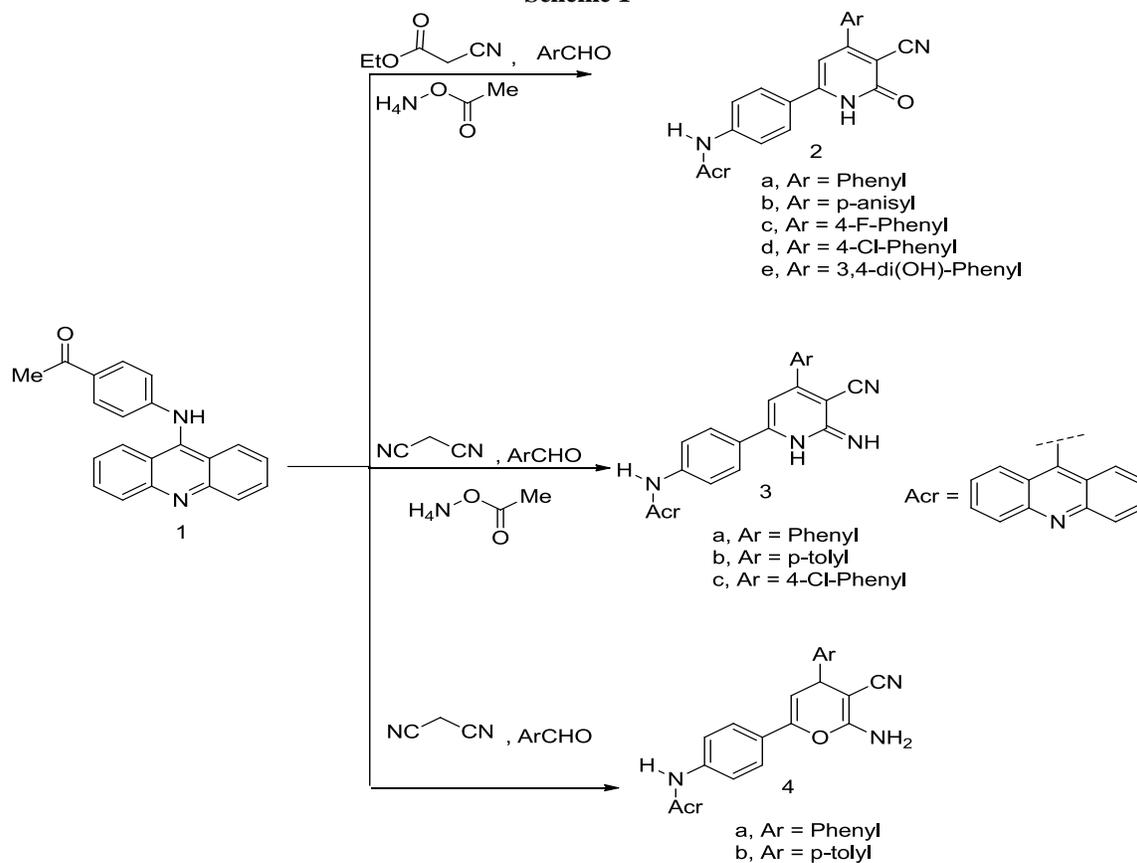
Figure 2. Molecular alignment between the gyrase enzyme of Bacillus subtilis (Uniprot code = P39814) and E-coli (uniprot code = P06612) with identity percentage = 33%

P39814	TOP1_BACSU	1	MSDYLVIIVESPAKAKTIERYLGGKYYKVKASMGHVRDLPKS-----	40
P06612	TOP1_ECOLI	1	MGKALVIIVESPAKAKTIEKYLGGSDYVYKSSVGHIRDLPTSGSAAKKSADSTSTKAKKPK	60
P39814	TOP1_BACSU	41	-----QMGVDIEQNFEPKYITIRGKGPVLKELKTAAKKAKKVYLAADPDREGEAIA	91
P06612	TOP1_ECOLI	61	KDERGALVNRMGVDPWHNWEAHYEVLPGEKVVSELKQLAEKADHIYLATDLDRGEAIA	120
P39814	TOP1_BACSU	92	WHLAHSLDLNLNSDCRVVFNEITKDAIKESFKHPRMINMDLVDAQARRILDRLVGYKIS	151
P06612	TOP1_ECOLI	121	WHLREVIGDDARYSRVVFNEITKNAIRQAFNKPGEELNIDRVNAQARRFMDRVVGYMVS	180
P39814	TOP1_BACSU	152	PILWKKVKKGLSAGRVQSVLRLLIDREKEINDFKPEEYWTIDGTFLLKQGE-TFEASFFG	210
P06612	TOP1_ECOLI	181	PLLWKKIARGLSAGRVQSVAVLVREREIKAFVPEEFWEVDASTTTPSGEALALQVTH	240
P39814	TOP1_BACSU	211	KNGKKLPLNSEADVKEILSQLKGNQYVTEKVKKERKRNLPFTTSTLQQEAARKLNFR	270
P06612	TOP1_ECOLI	241	QNDKPFPRVKNKEQQAQAVSLEKARYSVLEREDKPTTSSKPGAPFITSTLQQAASTRLLGFG	300
P39814	TOP1_BACSU	271	AKKTMIAAQLYEGIDLGREGTVGLIITYMRTDSTRISNTAVDEAAAFIDQTYGKEFLGGK	330
P06612	TOP1_ECOLI	301	VKKTMMIAQRLYE-----AGYIITYMRTDSTNLSQDAVNMVRGYSIDNFKKYLPEP	351
P39814	TOP1_BACSU	331	RKPAKKNENAQDAHEAIRPTSVLRKPSSELKAVLGRDQMRLYKLIWERFVASQMAPAVLDT	390
P06612	TOP1_ECOLI	352	PNQYASKENSQEAHEAIRPSDNNVMAESLKD-MEADAQLYQLIWRQFVACQMTPAKYVDS	410
P39814	TOP1_BACSU	391	MSVDLTNNGLTFRANGSKVVFSGFMKVYVEGKDDQMEEKDRMLPDLQEGDVLSKDIEPE	450
P06612	TOP1_ECOLI	411	TTLTVGAGDFRLKARGRILRFDGWTKVMPALR---KGDEDRILPAVNKGDALTLVELTPA	467
P39814	TOP1_BACSU	451	QHFTQPPRYTEARLVKLTLEERGIGRPSTYAPTLDTIQRRGYVALDNKRFVPTLGGIVL	510
P06612	TOP1_ECOLI	468	QHFTKPPARFSEASLVKELEKRGIGRPSTYASIIISTIQDRGYVVRNRRFYAEKNGEIVT	527
P39814	TOP1_BACSU	511	DLIMEFFPEIINVEFTAKMERDLDHVEEGNTEWVKIIDNFYTDKFRVKKAESEMKEVEI	570
P06612	TOP1_ECOLI	528	DRLEENFRELINNYDFTAQMENSLDQVANHEAENKAVLDHFFSDFTQLDKAEKDPPEEGM	587
P39814	TOP1_BACSU	571	EPE---YAGEDCELSSPMVYKMGRYGKFLACSNFPD----CRNFKPIV-----	612
P06612	TOP1_ECOLI	588	RPNQWLTSIDCPTCGRKMGIKRTASTGVFLGCSGYALPPKERCKTTINLVPENEVLNMLE	647
P39814	TOP1_BACSU	613	-----KQIGVVKPCSCGEGNIVERKSKKRVFYGCDRYPDCEFV-----SWDK	654
P06612	TOP1_ECOLI	648	GEDAETNALRAKRRCPKCGTAMDSYL-IDPKRKLHVCGNPNCTDGYEIEEGEFKRGYVDG	706
P39814	TOP1_BACSU	655	PIERKCPKCGKMLVEKLLKKGIVQVCEDYKEEPQK-----	691
P06612	TOP1_ECOLI	707	PI-VECEKCGSEMHLKMGFRGKYMACEEKNTRKILRNGEVAPPKEDPVPLPELPCEK	765
P39814	TOP1_BACSU	692	-----	691
P06612	TOP1_ECOLI	766	SDAYFVLRDGAAGVFLAANTFPKSRETRAPLVEELYFRDRLEPKLRYLADAPQQDPEGN	825
P39814	TOP1_BACSU	692	-----	691
P06612	TOP1_ECOLI	826	KTMRVFSRKTQQYVSSSEKDGKATGWSAFVVDGKWEVGGK	865

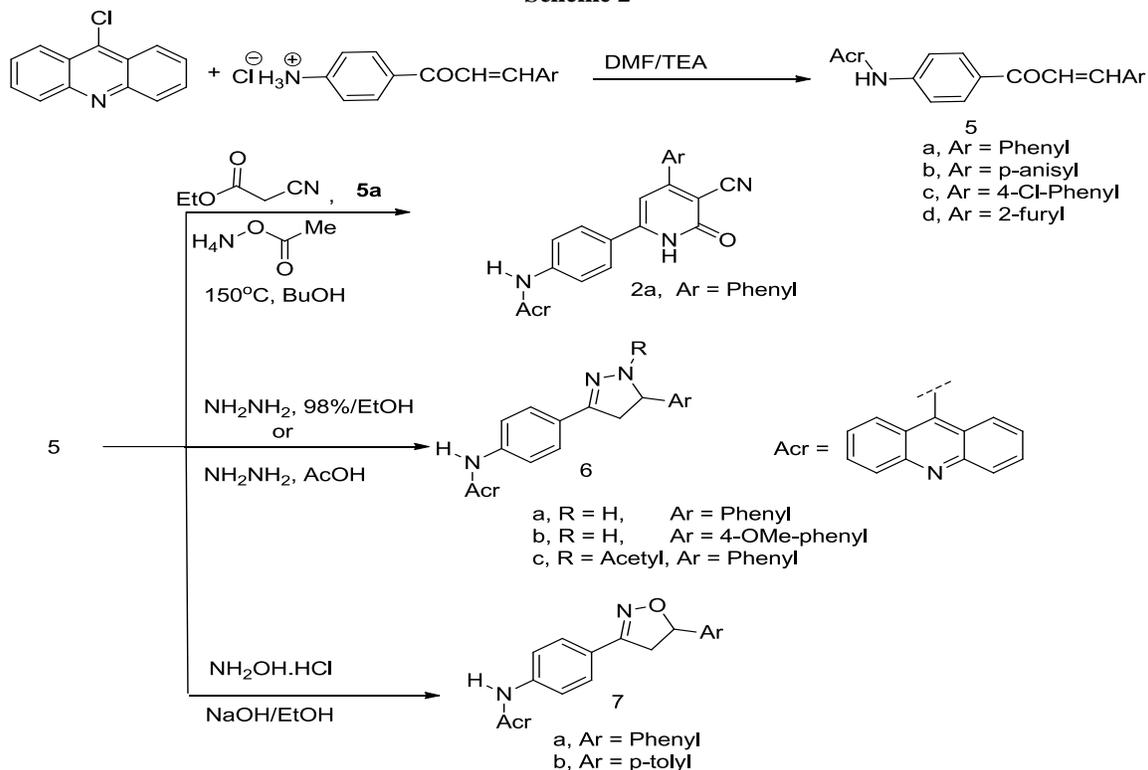
Figure 3. Molecular alignment between the gyrase enzyme of Bacillus subtilis (Uniprot code = P39814) and Staph. aerus (uniprot code = Q2FHI8) with identity percentage = 67.43%.

P39814	TOP1_BACSU	1	MSDYLVIIVESPAKAKTIERYLGGKYYKVKASMGHVRDLPKSQMGVDIEQNFEPKYITIRGK	60
Q2FHI8	TOP1_STAA3	1	MADNLVIIVESPAKAKTIEKYLGGKYYKVIASMGHVRDLPRSQMGVDTEDNYFEKYYITIRGK	60
P39814	TOP1_BACSU	61	GPVLKELKTAAKKAKKVYLAADPDREGEAIAWHLAHSLDLNLNSDCRVVFNEITKDAIKE	120
Q2FHI8	TOP1_STAA3	61	GPVVKELKHKAKKAKNVFLASDPDRGEAIAWHLASKILELDSKENRVVFNEITKDAVKE	120
P39814	TOP1_BACSU	121	SFKHPRMINMDLVDAQARRILDRLVGYKISPIWKKVKKGLSAGRVQSVLRLLIDREK	180
Q2FHI8	TOP1_STAA3	121	SFKNPREIEMNLVDAQARRILDRLVGYNISPLWKKVKKGLSAGRVQSVLRLLVLDREN	180
P39814	TOP1_BACSU	181	EINDFKPEEYWTIDGTFLLKQGETFEASFFGKNGKKLPLNSEADVKEILSQLKGNQYVTEK	240
Q2FHI8	TOP1_STAA3	181	EIRNFKPEEYWTIEGEFRYKSKSFNAKFLHYKKNPKLTKKDVKEITAAALDGDQFEITN	240
P39814	TOP1_BACSU	241	VTKKERKRNLPALPFTTSTLQQEAARKLNFRKAKTMMIAAQLYEGIDLGREGTVGLIITYMR	300
Q2FHI8	TOP1_STAA3	241	VTKKEKTRNPANPFTTSTLQQEAARKLNFRKARKTMMVAQQLYEGIDLKKGITIGLITYMR	300
P39814	TOP1_BACSU	301	TDSTRISNTAVDEAAAFIDQTYGKEFLGGKRKPAKKNENAQDAHEAIRPTSVLRKPSSELK	360
Q2FHI8	TOP1_STAA3	301	TDSTRISDTAKVEAKQYITDKYGESYTSKRKAS--GKQGDQAHEAIRPSSMTRTPDDMK	358
P39814	TOP1_BACSU	361	AVLGRDQMRLYKLIWERFVASQMAPAVLDTMSVDLTNNGLTFRANGSKVVFSGFMKVYVE	420
Q2FHI8	TOP1_STAA3	359	SFLTQDQYRLYKLIWERFVASQMAPAVLDTVSLDITQGDIKFRANGQTIKFKGFMTRYVE	418
P39814	TOP1_BACSU	421	GKDDQMEEKDRMLPDLQEGDVLTKDIEPEQHFTQPPRYTEARLVKLTLEELKIGRPSTY	480
Q2FHI8	TOP1_STAA3	419	TKDDSDSEKENLPLKLEQGDQVATQIEPAQHYTQPPRYTEARLVKLTLEELKIGRPSTY	478
P39814	TOP1_BACSU	481	APTLDTIQRRGYVALDNKRFVPTLGGIVLDLIMEFFPEIINVEFTAKMERDLDHVEEGN	540
Q2FHI8	TOP1_STAA3	479	APTIDTIQKRNYYKLESKRFVPTLGEIVHEQVKEYFPEIIDVEFTVNETLLDKIAEGD	538
P39814	TOP1_BACSU	541	TEWVKIIDNFYTDKFRVKKAESEMKEVEIEPEYAGEDCELSSPMVYKMGRYGKFLACSN	600
Q2FHI8	TOP1_STAA3	539	ITWRKVIDGFFSSFKQDVERAEEEMEIEIKDEPAGEDCEICGSPMVKMGRYGKFMACS	598
P39814	TOP1_BACSU	601	NFPDCRNTKPIVQKIGVVKPCSCGEGNIVERKSKKRVFYGCDRYPDCEFVSWDKPIERKC	660
Q2FHI8	TOP1_STAA3	599	NFPDCRNTKAIKVSIGVVKPCKNDGDVERKSKKRNRFYGCSDYKPECFISWDKPIGRDC	658
P39814	TOP1_BACSU	661	PKCGKMLVEKLLKKGIVQVCEDYKEEPQK-----	691
Q2FHI8	TOP1_STAA3	659	PKCNQYLVENKKGKTTQVICSNCDYKEAAQK	689

Scheme 1



Scheme 2



CONCLUSION

Several compounds with the 9- amino or anilino acridine core were prepared and their anti-microbial properties were studied. The antimicrobial potency of some the newly synthesized compounds were subjected for further docking studies to explore the binding pattern against *Bacillus Subtilis* (gram positive bacteria).

REFERENCES

1. Santelli-Rouviers, C, Barret J-M, Farrell CM, Sharples D, Hill BT, Barbs J. Synthesis of 9-acridinyl sulfur derivatives: sulphides, sulfoxides and sulfones. Comparison of their activity on tumor cells. *Eur. J. Med. Chem*, 39(12), 2004, 1029.
2. Chen K-M, Sun Y-W, Tang Y-W, Sun Z-Y, Kwon C-H. Synthesis and antitumor activity of sulfur containing 9-anilinoacridines. *Mol. Pharm*, 2(2), 2005, 118.
3. Dickens BF, Weglicki WB, Boehme PA, Mak TI. Antioxidant and lysosomotropic properties of acridine-propranolol. *J. of Mol. Cell. Cardiol*, 34(2), 2002, 129.
4. Chen YL, Lu CM, Tsao LT, Wang JP. Synthesis and anti-inflammatory evaluation of 9-anilino and 9-phenoxy acridine derivatives", *J. Med. Chem*, 45(21), 2002, 4689.
5. Kalirajan R, Muralidharan V, Jubie S, Sanker S. Microwave assisted synthesis, characterization and evaluation for their antimicrobial activities of some novel pyrazole substituted 9-anilinoacridine derivatives. *Int. J. Health and Allied Science*, 2(2), 2013, 81.
6. Gamage SA, Tepsiri N, Wilairat P, Wojcik SJ, Figgitt DP, Ralph RK, Denny WA. Synthesis and *in vitro* evaluation of 9-anilino-3,6-diaminoacridines active against a multi drug-resistant strain of the malaria parasite plasmodium Falciparum. *J. Med. Chem*, 37(10), 1994, 1486.
7. Zhang W, Zhang B, Zhang W, Yang T, Wang N, Gao C, Tan C, Liu H, Jiang Y. Synthesis and antiproliferative activity of 9-benzylamino-6-chloro-2-methoxy-acridine derivatives as potent DNA-binding ligands and topoisomerase II inhibitors. *Eur. J. of Med. Chem*, 116, 2016, 59.
8. Galdino-Pitta MR, Pitta MGR, Lima MCA, Galdino SL, Pitta IR. Niche for acridine derivatives in anticancer therapy. *MiniRev. Med. Chem*, 13, 2013, 1256.
9. Belmont P, Dorange I. Acridine/acridone: a simple scaffold with a wide range of application in oncology, *Expert. Opin. Ther. Pat*, 18, 2008, 1211.
10. Kaur J, Singh P. Acridine derivatives: a patent review (2009-2010), *Expert. Opin. Ther. Pat*, 21, 2011, 437
11. May BC, Witkop J, Sherrill J, Anderson MO, Madrid PB, Zorn JA, Prusiner SB, Cohen FE, Guy RK. Structure-activity relationship study of 9-aminoacridine compounds in scrapie-infected neuroblastoma cells. *Bioorg. Med. Chem. Lett*, 16, 2006, 4913.
12. Oxoby M, Moreau F, Durant L, Denis A, Genevard JM, Vongsouthi V, Escaich S, Gerusz V. Towards Gram-positive antivirulence drugs: new inhibitors of *Streptococcus agalactiae* Stk1, *Bioorg. Med. Chem. Lett*, 20, 2010, 3486.
13. Auparakkitanon S, Noonpakdee W, Ralph RK, Denny WA, Wilairat P. Antimalarial 9-anilinoacridine compounds directed at hematin. *Antimicrob. Agents Chemother*, 47(12), 2003, 3708.
14. Szmigiero L, Gniazdowski M. Complexes of nitracrine with DNA. Stoichiometry of binding. *Arzneimittelforschung*, 31(11), 1981, 1875.
15. Ledochowski A, Stefanska B. Nitracrine. *Rocz. Chem*, 40, 1966, 30; C. A. 1966, 65, 2219b.
16. Glaz TE, Talas M, Acranil. *Arch. Virol*, 48, 1975, 375.
17. Ferreira R, Aviñó A, Mazzini S, Eritja R. Synthesis, DNA-binding and antiproliferative properties of acridine and 5-methylacridine derivatives. *Molecules*, 17, 2012, 7067.
18. Palchadhuri R, Hergerother PJ. DNA as a target for anticancer compounds: Methods to determine the mode of binding and the mechanism of action. *Curr. Opin. Biotechnol*, 18, 2007, 497.
19. Neto BA, Lapis AA. Recent developments in the chemistry of deoxyribonucleic acid (DNA) intercalators: Principles, design, synthesis, applications and trends. *Molecules*, 14, 2009, 1725.
20. Safwat HM, Ragab FA, El-Said MK, El-Sayed NM. Synthesis, anti-tumor and antimicrobial activities of 3-chloro-9- (p-N-substituted sulfamoylphenylaminoethylene) acridines. *Egypt J.Pharm.Sci*. 33(3-4), 1991, 581-600.
21. Rasotgi K, Chang JY, Pen WY, Chen CH, Chou TC, Chen LT, Su TL. Antitumor AHMA linked to DNA minor groove binding agents, synthesis and biological evaluation. *J. Med. Chem*, 45(20), 2002, 4485.
22. Makhlof AA, Kamel MM, Anwar MH, Haiba ME, Mohei El-Deen EM. Synthesis of some new 1,8-naphthyridine-3-carboxamides of possible antimicrobial activity. *Egypt Pharm. J*, 4(2), 2005, 371.
23. Auclair C, Voisin E, Banoun H, Paoletti C, Bernadou J, Meunier B, Potential antitumor agents: synthesis and biological properties of aliphatic amino acid 9- hydroxyellipticinium derivatives. *J. Med. Chem*, 27(9), 1984, 1161.

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CONFLICT OF INTEREST

The authors declare that they have no conflict of interest.

24. Albert A. The Acridines, 2nd Ed., *Edward Arnold Ltd., London*, 1966, 519.
25. Mosher MD, Holmes KL, Frost K.S. Structure-Activity Relationships for the 9-(Pyridin-2'-yl)-aminoacridines. *Molecule* 2004, 9, 102.
26. El-Taliawy GM, Ali EI, Hegazy GH, Ismail NS, Ramadan W. Computer aided drug design, synthesis and biological evaluation of novel acridine derivatives as topoisomerase I inhibitors. *J. Amer. Sci*, 6(11), 2010, 148.
27. Kamel M, Nabih I, Gadalla KZ, Ashour MM. New pyrazolinylaminoquinolines and other related products of possible antibacterial activity. *Die Pharmazie*, 41(1), 1986, 55.
28. Kamel MM, Kassem EMM, Fahmy HH, Abdou WAM. Synthesis and preliminary antimicrobial screening of some new tetrahydronaphthyl heterocycles. *Egypt J. Chem*, 39(3), 1996, 271.
29. Kamel MM, Omar MT, Refai M, Fahmy HH, Nofal ZM, Ismail NS. Synthesis and antimicrobial evaluation of some new 1,8-Naphthyridine derivatives. *Egypt J. Chem*, 39(6), 1996, 591.
30. Kamel MM, Omar MT, Kassem EMM, Khalifa NM. New 4-substituted phenoxyquinolines of possible antimicrobial activity. *Egypt J. Pharm. Sci*, 38(1-3), 1997, 61.
31. Kamel MM, Nofal ZM, Fahmy HH, Refai M, Haiba ME. New quinolines and thiazoloquinolines with possible antitumor activity. *Proc. Pakistan Acad. Sci*, 37(1), 2000, 41.
32. Vitalino de Almeida SM, Lafayette EA, Gomes da Silva LPB, da CA, Amorim C, Bento de Oliveira T, Gois Ruiz ALT, Ernesto de Carvalho J, Olímpio de Moura R, Carneiro Beltrão EI, Alves de Lima M, Júnior LB de C. Synthesis, DNA binding, and antiproliferative activity of novel acridine-thiosemicarbazone derivatives. *Int. J. Mol. Sci*, 16, 2015, 13023.
33. Chen YL, Lu CM, Chen IL, Tsao LT, Wang JP. Synthesis and antiinflammatory evaluation of 9-anilino acridine and 9-phenoxy acridine derivatives. *J. Med. Chem*, 45(21), 2002, 4689.
34. Attia A, Michael M. Azachalcones. I. Synthesis and antimicrobial activity of newer cyanopyridines, *Acta Chim. Hung*, 112, 1983, 89.
35. Benny WA, Cain BF, Atwell DJ, Hansch C, Panthanackal A, Leo A. Potential antitumor agents. Quantitative relationships between experimental antitumor activity, toxicity, and structure for the general class of 9-anilinoacridine antitumor agents. *J. Med. Chem*, 25(3), 1982, 276.
36. Nezval J, Halacka K. The enhancing effect of EDTA on the antibacterial activity of nalidixic acid against *Pseudomonas aeruginosa*. *Experientia*, 23(12), 1967, 1043.
37. Shadomy S. *In Vitro* Antifungal Activity of Clotrimazole (Bay b 5097). *Infect. Immun*, 1971, 143
38. Abou-Zeid AA, Shehata YM, A simple technique for assaying antibiotics using methylene blue as an indicator. *Indian J. Pharm*, 31, 1969, 72.
39. Molecular Operating Environment (MOE), 2013.08; Chemical Computing Group Inc., 1010 Sherbooke St. West, Suite #910, Montreal, QC, Canada, H3A 2R7, 2016.
40. Trott O, Olson AJ. Auto Dock Vina: improving the speed and accuracy of docking with a new scoring function, efficient optimization, and multithreading. *J. Comput. Chem*, 31(2), 2010, 455.
41. Benoit AR, Schiaffo C, Salomon CE, Goodell JR, Hiasa H, Ferguson DM. Synthesis and evaluation of N-alkyl-9-amino acridines with antibacterial activity. *Bioorg. & Med. Chem. Lett*, 24(14), 2014, 3014.
42. Prescott TA, Sadler IH, Kiapranis R, Maciver SK. Lunacridine from *Lunasia amara* is a DNA intercalating topoisomerase II inhibitor. *J. Ethnopharmacol*, 109(2), 2007, 289.
43. Sanner MF. Python: a programming language for software integration and development. *J. Mol. Graph. Model*, 17(1), 1999, 57.